Purification of recombinant Gcn4

James Fishburn, Hahn Lab March 2006

Gcn4 constructs are cloned into the pET21(a) vector and have a 6-His tag expressed at the N-terminus

Gcn4 is expressed and purified from BL21(DE3)RIL cells

Day 1

Grow 10+ ml culture overnight

Day 2

- 1. Inoculate 1 L media with 10 ml saturated overnight
- 2. Grow cells to OD600 0.6 (ca. 3hrs at 37 deg)
- 3. Induce cells with 1 mM IPTG
- 4. Continue incubation for 3 hrs at 37 deg with shaking
- 5. Harvest cells- 5' spin at 5000 rpm in GSA
- 6. Wash cells with 50 ml Binding buffer
- 7. Resuspend cells in 20 ml Binding buffer
- 8. Freeze and store at -70 deg

Day 3

- 1. Thaw cells
- 2. Lyse cells with 1 pass through microfluidizer
- 3. Clarify extract by centrifugation- 20' spin at 10,000rpm in SS34
- 4. Transfer clarified extract to 50 ml tube
- 5. During spin prepare Ni-Sepharose: wash with 5 volumes DI water, wash with 5 volumes Binding buffer, resuspend Ni-Sepharose to 50% in Binding buffer (spins are at 500 x g for 2'-5')
- 6. Add 2 ml 50% Ni-Sepharose slurry to extract
- 7. Incubate extract with resin for 30' at RT with mixing (nutate)
- 8. Collect resin by centrifugation, remove supernatant and save (flow through)
- 9. Wash resin a total of three times using 5 volumes of Binding buffer for each wash
- 10. Add 2 volumes Elution buffer to resin and incubate 5' at RT with mixing (nutate)
- 11. Collect resin and save first elution
- 12. Repeat elution step 2-4 more times saving each elution

- 13. Analyze purification by SDS-PAGE (4-12% Bis-Tris, MES) and CBB staining- 2 microliter of each elution per lane is sufficient for gel staining along with 1microliter each of crude lysate, clarified lysate, and the flow through
- 14. Combine desired elutions, freeze, and store protein at -70 deg Day 4
- 1. Thaw protein
- 2. Dilute Gcn4 to be purified 1:5 in 20 mM sodium phosphate pH 7.0 to bring NaCl concentration to 100 mM
- 3. Filter sample to be purified through 0.45 micron syringe filter disc
- 4. Prepare AKTA FPLC by established protocols (filter buffers)
- 5. Prepares BioRex70 column by washing first with water, then BioRex70 buffer B, and finally with 10% BioRex70 buffer B until conductivity and UV are stable
- 6. Prepare fraction collector as desired
- 7. Load Gcn4 onto FPLC
- 8. Run program "BioRexmGcn4NewSystem": Unicorn software, user = JamesF, password = default, program was created 8/18/05 and last modified 9/2/05

Notes

- Capacity of BioRex70 appears to be 3-4 mg of Ni-Sepharose purified Gcn4
- Gcn4 will elute from BioRex70 at approximately 0.2 mg/ml
- 1 mM PMSF and 3 mM DTT are added to all buffers for Ni-Sepharose purification on days 2 and 3
- Using protease tablets instead of PMSF does not increase Gcn4 stability during lysis and Ni-Sepharose purification
- 0.5 mM PMSF and 5 mM DTT are added to buffers for BioRex70 purification on day 4

Buffers

Binding buffer: 20 mM sodium phosphate, 0.5 M NaCl, 20 mM imidazole, pH 7.4

Elution buffer: 20 mM sodium phosphate, 0.5 M NaCl, 500 mM imidazole, pH 7.4

BioRex70 buffer A: 20 mM HEPES, 1 mM EDTA, 10% glycerol

BioRex70 buffer B: 20 mM HEPES, 1 mM EDTA, 10% glycerol, 1 M KCl