## Acetone precipitation of protein

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This procedure is suitable for recovering proteins from most aqueous solvents and from SDS containing buffers. It is not recommended for proteins dissolved in urea or quanidine or for peptides.

## **Protocol**

- 1. Cool the required volume of acetone to -20°C.
- 2. Place protein sample in acetone-compatible tube, such as polypropylene and able to hold six times the sample. Screw cap tubes may help minimize sample losses.
- 3. Add six times the sample volume of cold (-20°C) acetone to the tube.
- 4. Vortex tube and incubate for 2 hours to overnight minutes at -20°C.
- 5. Centrifuge 15 minutes at 13,000-15,000 x g at 4°C.
- 6. Decant and properly dispose of the supernatant, being careful to not dislodge the protein pellet.
- 7. Briefly wash the pellet with 100ul of cold 90% acetone.
- 8. Centrifuge 5 minutes at 13,000-15,000 x g at 4°C.
- 9. Remove sup and repeat if necessary.
- 10. Air dry for ~15-30 minutes and resuspend in an appropriate buffer.