

## Expand High Fidelity PCR (Boehringer)

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Expand is a mixture of Taq and Pwo DNA polymerases (Boehringer). Pwo polymerase has 3' to 5' exo proofreading activity. If amplifying a 1 Kb fragment for 20 cycles, expect ~8% of the products will have at least one mutation. See Boehringer data sheet for more details on error rate.

Mix in an 0.2 ml thin wall microfuge tube:

- 1 microliter each: dATP, dGTP, dCTP, dTTP (10 mM solutions)
- 100 ng each primer (20-mer)
- 5 microliters Expand buffer +15 mM MgCl<sub>2</sub>
- DNA template (typically ~15 ng of plasmid (1 microliter of a 1/15 dilution of a miniprep), or 50-100 ng yeast genomic DNA (1-2 microliters of a 1/10 dilution of yeast miniprep DNA))
- H<sub>2</sub>O to a final volume of 50 microliters
- 0.75 microliters Expand enzyme

Use the following profile in the temp. cycler:

1. 94 degrees 2 min.
2. 94 degrees 20 sec.
3. 45-65 degrees 30 sec. (temp. will depend on annealing temp. of primers)
4. 72 degrees (<3 Kb) or 68 degrees (>3 Kb) for 0.7 min x Kb of product

repeat steps 2-4 for 15-25 times (15 is probably plenty for amplifying DNA from plasmid).

If doing more than 15 cycles, elongate each additional cycle by 5 sec. to compensate for reduced activity of the polymerase.

Hold at 4 degrees when finished.

If adding a sequence to a PCR product such as a His tag, etc it is sometimes a good idea to do the first 10 cycles at a lower annealing temperature consistent with the exact homology of the primers to the

DNA target and then raise the annealing temperature for the next 15 cycles to take into account the longer region of homology.

Note: keeping the extension time to the recommended time will help to reduce the error rate. When Polymerase misincorporates a base, it is slow to put in the next base. Therefore, extended elongation times will allow the polymerase to get past the base misincorporation.